

**Diagnostics Navicent Health, Laboratory**  
**Department of Pathology and Laboratory Medicine**  
Laboratory Collection Information

## TABLE OF CONTENTS

	PAGE
<b>General Information</b>	<b>2</b>
General Laboratory Hours	2
Supplies	2
Courier Service	3
Billing Information	3
Billing Guidelines	4
<b>Specimen Collection</b>	<b>6</b>
Blood Collection Tubes	6
Specimen Identification	6
Minimum Volumes	6
Specimen Stability	6
CLSI (NCCLS) order of draw	7
Urine Collection	7
Urine 24 Hour Collection	7
Body Fluids	7
Criteria for Specimen Rejection	7
Bacteriological Specimen Collection	8
Anatomic Pathology and Cytology Specimen Collection	13
Test Profiles	17

Rev: February 17, 2023  
Reviewed:

# GENERAL INFORMATION

## GENERAL LABORATORY HOURS

The laboratory is open 24-hour basis. Outpatient Lab Collection hours are listed below.

### Patient Service Centers

#### **Diagnostics Hardeman Navicent Health**

1650 Hardeman Ave.

Macon, Ga. 31201

478-633-1234

Fax: 478-633-5200

Hours: Monday- Thursday

8:00 a.m. – 6:00 p.m.

Hours: Friday

8:00 a.m. – 5:30 p.m.

#### **Diagnostics Zebulon Road Navicent Health**

5925 Zebulon Rd.

Macon, Ga. 31210

478-757-7877

Fax: 478-757-7876

Hours: Monday – Friday 8:00 a.m. – 5:00p.m.

#### **Diagnostics Monroe Navicent Health**

120 North Lee Street

Forsyth Ga 31029

478-993-9900

Fax: 478-993-9907

Hours: Monday- Friday 8:00 am to 4:30 pm

#### **Diagnostics Pine Street Navicent Health**

770 Pine Street

Macon Ga 31201

478-633-0034

Fax: 478-633-5200

Hours: Monday – Friday 8:00 a.m. – 5:00p.m.

### **Diagnostics Navicent Health Lab Services Telephone Numbers**

Manager Outpatient Laboratory (478) 633-1072

Clinical Laboratory Supervisor (478) 633-2092

Clinical Laboratory Supervisor Offsite (478) 633-2646

Client Service Representative (478) 633-1234

Laboratory Front Desk (478) 633-1234

Toll Free (888) LAB-XRAY

## SUPPLIES

Biohazard transport bags, containers for urine and stool specimens, blood drawing supplies, and request forms are provided to outreach clients at no charge. Special collection containers are also provided. Please contact the Diagnostics Navicent Health, Laboratory Client Services 478-633-1234.

## COURIER SERVICE

Scheduled and stat courier service is provided Monday- Sunday. For information on this service, please contact the Diagnostics Navicent Health, Laboratory Client Service Representative 478-633-1234.

## RESULTS REPORTING

Individual patient results include the patient's name, lab identification number, collection times, date, result, and normal values. All results are faxed or transmitted electronically to the ordering physician. All critical values are called immediately to the provider. Other results are called upon request during normal office hours.

## BILLING INFORMATION

Several billing options are available to Diagnostics Navicent Health, Laboratory clients. You may elect client billing, third party billing or patient direct billing.

**Client:** An itemized invoice will be sent monthly to the referring institution, health care provider, physician, group practice. This invoice will include a list of charges for tests performed on each patient. Payment is due within 30 days of receipt of the invoice.

Proper laboratory requisitions must be sent with patient or samples to ensure correct billing for client accounts. The requisition must be marked Bill to: Account. Client accounts will be registered in the Atrium Health business office and must be approved by the Compliance and Contracting.

### 3<sup>rd</sup> Party/Medicare/Medicaid:

For Atrium Health to properly bill for lab services, we depend on you to provide us with your Patient's **complete** billing information, including all medically appropriate diagnosis ICD-10 codes. If this information is not included on your patient's original requisition for lab services, then we must contact your office to obtain this information. If all billing information is not received your patient may receive a bill.

The following is a list of what is considered **complete** billing information:

- \* Bill to: account, patient, other
- \* Patient name (last, first, middle)
- \* Date of birth
- \* Sex
- \* Patient Social Security #
- \* Patient phone #
- \* Print name of insured/resp. party (last, first, middle) if other than pt.
- \* Street address of insured/resp. party
- \* Medicare / Medicaid number
- \* Relationship to insured
- \* Insurance company name
- \* Member/insured ID#
- \* Group #
- \* Insurance address
- \* ICD10 diagnosis code(s) for tests ordered
- \* Tests ordered on Medicare patients must be reviewed for Medical Necessity

\*\* A copy of the insurance card front and back may be attached to the lab requisition form.

**Patient Billing- (Self Pay):** Bill is sent directly to patient/guarantor at the home address. No insurance or third party is filed.

If you elect to have your patient billed by Atrium Health, the patient becomes responsible for the full payment of the bill. If the patient has insurance coverage, please refer to the section of this manual that describes Third Party Billing information requirements. When you select to bill the patient directly, the following information:

- Patient's Name
- Current Address (Including Apt. #)
- Zip Code

- Patient Social Security Number
- Date of birth
- Patient Telephone Number (Including Area code)

must appear clearly on the test requisition, in the areas provided. Provide all requested information to avoid follow-up correspondence to you from the billing Department. Please be sure to inform your patient that they will be receiving a laboratory bill from NAVICENT HEALTH. Patient payments are due by the date indicated in the billing information section, which is located in the upper right hand corner of the bill.

## Payments

Payments may be mailed to:

Medical Center Navicent Health  
P.O. Box 116417  
Atlanta Ga 30368-6417

Billing Inquiries:

Billing Customer Service : 478-633-3200

## Third Party Billing Information

Diagnostics Navicent Health, Laboratory services can and will accept payment from many agencies and insurance companies. To avoid follow-up correspondence from the Billing Department, provide all the billing information, including all medically appropriate current ICD10 diagnostic codes, on the requisition in the areas provided.

Review your patient's insurance identification cards to ensure accurate Identification Number, correct spelling of names as they appear clearly on the requisition. **Patients will be responsible for charges such as non-covered services, co-pay amounts, deductible amounts, etc. depending upon the type of coverage they have.**

Please communicate this information to your patient at the time of service.

## Billing Guidelines

Payors (Including Medicare) follow medical necessity guidelines. All payors require laboratories to include diagnoses (diagnosis, symptom, condition, complaint or problem) on every claim as documentation of medical necessity.

Diagnosis information must be furnished using the ICD-10-CM coding system. Health Care Financing Administration (HCFA) guidelines indicate that the ultimate responsibility for providing correct ICD-10 codes lies with the ordering physician. All Medicare carriers have implemented Local Medical Review Policies that restrict the medical conditions under which Medicare payment for certain tests will be made. These policies define the medical conditions (diagnosis, symptom, condition, complaint or problem) or ICD-10 codes that establish the medical necessity of certain tests under which payment will be made. If a limited coverage test is ordered in conjunction with a diagnosis code that is not included in the Medicare carrier's predetermined list of diagnoses, Medicare will not pay for the test.

Any diagnosis information submitted on Laboratory claims must be furnished by the ordering physician or his or her authorized staff and must be consistent with the diagnosis information documented in the patient's medical records for that date of service.

Additionally, we need you to provide an **Advance Beneficiary Notice (ABN) (formerly known as Patient Acknowledgement of Non-Covered Services [PANS])** form when a limited coverage tests is ordered and the diagnosis provided does not support medical necessity (making the test likely to be

denied by Medicare). The ABN form should be signed by the patient for each test requisition on which a limited coverage test is ordered, either individually or as part of a panel, when the diagnosis does not support medical necessity. This will allow NAVICENT HEALTH to invoice the patient for the test if it is not covered within the diagnosis restrictions in the Local Medical Review Policies. The laboratory has modified its test requisition to include test CPT codes and to provide room for more diagnosis codes. The ABN form should be completed by you and the patient and submitted along with the test requisition, that includes an order for any non-covered tests, as well as tests not covered by Medicare because they are considered to be experimental or used primarily for screening purposes, (for example, tests that have not yet been approved by the FDA are excluded from Medicare coverage).

**The Medicare Website is available for you to use as a quick reference tool in determining whether a particular limited coverage test is covered for your Medicare patient's diagnosis.** <https://www.cms.gov/medicare-coverage-database/overview-and-quick-search.aspx> The ICD-10 codes provided on the test requisition (which will in turn be submitted to Medicare and other payors) MUST be consistent with the information contained in the patient's medical records for that date of service. The ICD-10 manual is the definitive resource for a complete listing of all ICD-10 codes.

Payors' recent medical necessity guidelines and Local Medical Review Policies represent a fundamental shift in the way in which physicians must document orders for clinical laboratory tests for Medicare patients, in summary:

1. If a limited coverage test is ordered for a patient under a medical condition (ICD-10 code) that the Medicare carrier deems as medically necessary, the laboratory must receive the ICD-10 code on the test requisition from the physician so that we may submit the ICD-10 code on the Medicare claim.
2. If a limited coverage test is ordered for a patient under a medical condition (ICD-10) that the Medical carrier deems as medically unnecessary and is likely to deny, the laboratory must receive the appropriate ICD-10 code that justifies the service, in the physician's judgement, **AND** an ABN form signed by the Medicare patient. (A properly executed ABN form is required to seek reimbursement directly from the Medicare patient if Medicare denies the claim.)

### **CPT Codes (Current Procedure Terminology Codes)**

**The indicated CPT Codes are provided for informational purposes only. This coding is based on our current test methodology. We cannot accept responsibility for reimbursement you may or may not receive based on the procedure coding we have provided. Any questions regarding program coding should be confirmed with the payor being billed.**

CPT codes are provided with each test for reference, in an effort to improve understanding of the diagnostic procedures performed. The application of a specific code to a given procedure is a matter of individual interpretation.

## **SPECIMEN COLLECTION**

### **BLOOD COLLECTION TUBES**

The following abbreviations for the appropriate Vacutainer blood collection tubes are in the test information found under individual test listings.

Gold	- gold top, 6 ml clot activated, gel, serum tube
Red	- small red stopper, 7 mL capacity, no additive
Blue	- blue stopper, 5 mL capacity, Na citrate 3.2 % Buffered
Lav	- lavender stopper, 3 mL or 5 mL capacity EDTA

- Pink - pink stopper, 6 mL EDTA Blood Bank
- Green - green stopper, 7 mL capacity, lithium heparin
- Dark Green - green stopper, 10 mL capacity, Sodium heparin
- BCul - blood culture bottle, collect sterile conditions
- Gray - gray stopper, 7mL capacity, Na fluoride

### Microvolume Collections

- Serum capillary tube (red cap): 0.5 mL capacity typically yields 0.20 mL of serum
- Heparin, lithium capillary tube (green top): 0.5 mL capacity yield 0.2 mL plasma
- EDTA capillary tube (lavender top): 0.25 mL minimum volume
- Pediatric blue top: 1.8 mL minimum volume
- Pediatric gray top (no fluoride/K oxalate): 0.250 mL

### SAMPLE IDENTIFICATION

Specimens must be clearly labeled with two unique identifiers, Patient name (Last, First), Date of Birth and/or lab requisition number. Specimens for testing by Blood Bank Department require Date and time, patient social security or Medical Record number, and phlebotomist initials. The labeling must match accompanying requisition. Specimens with no identification or incorrect information will be rejected by the laboratory. The client will be notified during business hours, laboratory client services will request recollection of sample.

### MINIMUM VOLUMES

When minimum volumes are listed, they represent the true minimums required to perform the test. These do not usually allow for repeat or confirmation testing. Collection of double the minimum is recommended whenever possible. **Please call the laboratory about specific test requirements for unlisted tests or any additional information you require.**

### SPECIMEN STABILITY

Specimens collected in gel barrier tubes (gold) must be centrifuged prior to delivery to lab. Samples with special handling are noted in the Diagnostics Navicent Health Test List, which is available on the Navicent Health Epic EMR and the external laboratory website at [www.navicenthealth.org](http://www.navicenthealth.org).

### CLSI (NCCLS) order of draw

1. Blood culture tubes (yellow stopper), or culture bottles
2. Citrate (light blue stopper), or coagulation tubes 3
3. Non additive, serum tubes (red stopper)
4. Gel separator tube (gold top), or SST
5. Heparin (green top)
6. Ethylene diamine tetra acetic acid (EDTA) (lavender top)
7. Blood Bank (EDTA) (Pink Top)
8. Oxalate/Fluoride (Gray Top)
9. Other Additive Tubes

### URINE COLLECTION CONTAINERS

Routine and 24-hour urine containers (with appropriate preservative) are available from the laboratory for Diagnostics Navicent Health, Laboratory clients.

## **COLLECTION OF 24-HOUR URINE SPECIMENS**

Unless the physician specifies otherwise:

- At specified time the patient empties bladder and specimen is discarded.
- All urine produced during the next 24 hours is saved and combined in the large plastic container.
- At the same time of the following day, the patient empties bladder and this urine is included in the pooled specimen.
- Specimen should be kept refrigerated or on ice during collection period and sent promptly to the laboratory.
- Some tests require preservatives, information regarding special handling and storage is provided to patient upon receipt of the container.

## **COLLECTION OF BODY FLUIDS**

Body fluids (ascitic, pleural, pericardial, etc) should be collected in a sterile container of appropriate size and sent promptly to the laboratory. Specimen collected in syringe must have needle removed prior to transport to lab.

## **CRITERIA FOR SPECIMEN REJECTION**

Below are general criteria which apply to all specimens. Other specific criteria which may apply to individual procedures are detailed in the causes for rejection section under the individual test listings.

- Specimen improperly labeled
- Specimen improperly collected and/or preserved
- Hemolyzed specimens
- Specimen sample volume insufficient for test(s) ordered (physician will be contacted to determine what tests he/she would like deleted because of short sample, except for coagulation tests which require the correct volume for any testing.)
- Outside of container contaminated with specimen, specimen leaked from container.
- Patient not properly prepared
- Blood Bank samples that are not labeled with name, SSN, date of collection, time of collection, and collector's initials

## **BACTERIOLOGICAL SPECIMEN COLLECTION---CULTURES**

### **Cultures**

#### **I. Aerobic and Anaerobic Cultures**

- A. Transport containers are available for Diagnostics Atrium Health, Laboratory clients.
  1. Swab specimens --- eSwab Copan Inc. for Aerobic, Anaerobic and Fastidious bacteria.
  2. Fluids or aspirates --- sterile container
- B. Unacceptable specimens for anaerobic culture include throat or n/p swabs, sputum, feces, urine (except for suprapubic aspirates), eye or ear specimens, and vaginal or cervical swabs not collected by visualization via speculum.

#### **II. Blood Cultures**

Clean the venipuncture site using the blood culture prep kit available from the laboratory. Instructions are included with the kit.

- A. If multiple tests are ordered, the blood culture sample is collected first.
- B. The rubber diaphragm tops of the blood culture bottle must be cleaned with alcohol before injecting blood.
- C. Routine blood cultures (available from lab): The type and number of blood culture bottles to use per venipuncture depends on the amount of blood collected. The optimum volume of blood is 20 mL. Use the chart below for determining what bottles should be used.

**Adults:**

<b>Blood Volume</b>	<b>BactAlert Aerobic (SA or FA) (Blue or Green Top)</b>	<b>BactAlert Anaerobic (SN or FN) (Burgandy or Orange Top)</b>
10-20mL	½ collected	½ collected
< 10 mL	All	None

**Pediatrics:**

<b>Blood Volume</b>	<b>BactAlert Aerobic (PF) (Yellow Top)</b>
1-5 mL	All

- D. AFB(TB) blood cultures (available from lab, Sodium Heparin (green top) --- using blood culture prep kit, collect 3-5 mL of blood.
- E. Fungal blood cultures (available from lab, Blue or GreenTop bottle) --- using blood culture prep kit, collect 5-10mL of blood.

**III. Endocervical Cultures**

- A. Obtain under direct vision with a speculum.
- B. Gonorrhoea culture or DNA probe
  - Culture – Cervix is wiped clean and fresh exudate is collected with a Eswab and or Culturette device. Swab is placed in container.
  - DNA probe ---Cervical swab-
    1. Special collection kits available from lab
    2. The cervix is wiped clean and a swab rolled firmly against the cervical wall in order to collect the epithelial cells.
    3. Urine- Collect in sterile urine container, first 15-20 ml of urine obtained after waiting at least 1 hour from prior urination. Refrigerate sample immediately and transport to lab on ice. (Ice packs available from lab on request.)
- C. Herpes simplex culture
  1. Special collection kits are required and are available from the laboratory.
  2. Fluid from lesions is collected on a swab and the swab is placed in a special tube of transport broth. (Viral/Chlamydia Media)
  3. The specimen is sent promptly to the lab or refrigerated for no longer than overnight.
- D. *Chlamydia* culture ---Cervical swab
  1. Special collection kits are available from the lab. (Viral/Chlamydia Media)
  2. The cervix is wiped clean and a swab rolled firmly against the cervical wall in order to collect the epithelial cells.
- E. *Chlamydia* DNA probe ---Cervical swab
  1. Special collection kits are available from the lab.



2. The cervix is wiped clean and a swab rolled firmly against the cervical wall in order to collect the epithelial cells.
3. Urine- Collect in sterile urine container, first 15-20 ml of urine obtained after waiting at least 1 hour from prior urination. Refrigerate sample immediately and transport to lab

#### **IV. RSV Direct Test (antigen detection)**

- A. Nasal aspirates
  1. Attach a No. 8 French soft plastic feeding tube through a valve-containing trap to an electric suction apparatus.
  2. Attach a sterile catheter tip to the tube and introduce the tip through the nares to the back of the nose.
  3. Apply suction intermittently while catheter is slowly withdrawn. Collect in a sterile container.
  4. Optimum specimen is 0.3 mL and deliver specimen to lab promptly.
- B. Nasal washings
  1. The point of a suction bulb containing 3-7 mL of saline is placed in the nose so as to completely occlude one side.
  2. The saline is then squeezed into the nose and rapidly aspirated.
  3. The secretions are expelled into a sterile container and promptly delivered to the lab.
- C. Nasopharyngeal swab
  1. A special nasopharyngeal swab (HydraFlock Sterile Flocked Collection Device) obtained from the laboratory is used to collect the specimen. The swab is inserted through the nose to the posterior nasopharynx, allowed to remain a few seconds then removed.
  2. An alternative method is to bend the swab near the tip and insert it through the mouth and behind the uvula and soft palate into the nasopharynx. Care should be taken to avoid oral cavity contamination.
  3. Place swab into container.
  4. Label and submit to laboratory

#### **V. Fecal (stool) Cultures and Parasitology Tests**

1. Specimens are to be collected in a clean, dry container free of urine and toilet bowl water.
2. For culture, place a tablespoon amount of formed stool or approximately 5 mL of liquid stool into a feces collection vial containing Carey-Blair medium (red colored). Shake to homogenize specimen.
3. For comprehensive ova and parasite microscopic examination, place a tablespoon amount of formed stool or approximately 5 mL of liquid stool into a vial containing SAF fixative (clear colored). Shake fixative vial to homogenize specimen.
4. For Giardia and/or Cryptosporidium antigen, a tablespoon portion or a few milliliters (if liquid) of feces is placed in an empty fecal transport container and/or a vial containing SAF fixative (clear colored).
5. For Gastrointestinal Panel a tablespoon portion or a few milliliters (if liquid) of feces is placed in an empty fecal transport container
6. Liquid stools for the examination of amebic trophozoites should be sent promptly to lab.
7. Specimens containing oils, bismuth, and barium are generally unsuitable for examination.

8. Specimens for Clostridium Difficile Toxin will be rejected if formed stool is submitted.

#### **VI. Fungus Cultures** (All specimens placed in sterile container)

Skin scrapings --- the infected area should be washed with 70% alcohol and scrapings should be taken from active border areas of lesion.

Nail scrapings --- the area of infection should be washed with 70% alcohol. The scrapings or clipping of the nail should be selected from areas of active infection.

Hair --- the basal portion of the hair or hair stubs may be submitted. Select hairs from the edges of infected areas.

Subcutaneous mycoses --- specimens include crusts, pus, exudates, aspirated fluid and tissue.

Systemic mycosis --- specimens include CSF, sputum, bone marrow, and tissue.

#### **VIII. Nasopharyngeal Cultures**

A nasopharyngeal swab is used to collect the specimen. The swab is inserted through the nose to the posterior nasopharynx, allowed to remain a few seconds then removed.

An alternative method is to bend the swab near the tip and insert it through the mouth and behind the uvula and soft palate into the nasopharynx. Care should be taken to avoid oral cavity contamination.

#### **IX. Sputum Cultures**

A sputum collection container is used provided to Laboratory clients.

The mouth should be rinsed and if dentures are present, they should be removed.

The specimen should represent a true pulmonary secretion after deep coughing. The first early AM specimen is preferable.

#### **X. Throat Cultures**

With the patient's tongue depressed and the throat well exposed, rub the swab over the back of the throat. Rub over areas of inflammation, exudation, or ulceration.

Follow directions on Culturette/Eswab package --- provided to lab clients.

#### **I. Urethral Cultures**

Specimens should not be collected until 1 hour after urinating. Culturette/Eswab provided to lab clients.

See Endocervical cultures for collection kits for Gonorrhea, Herpes, and Chlamydia testing.

#### **XI. Urine Cultures**

- A. A catheterized, clean-catch, midstream, suprapubic aspiration or urine from an indwelling catheter may be submitted with the method of collection stated on the requisition.
- B. When an indwelling catheter is in place, urine should not be collected from the drainage bag.
- C. In the clean-catch midstream technique, the periurethral area is carefully cleansed with towelettes or Soap-soaked sponges and well rinsed with warm water to remove the detergent. The first portion of the urine is discarded and the subsequent portion is passed into an acceptable container.
- D. A 24-hour collection for AFB culture is not acceptable because of urethral contamination and the dilution of the acid fast organism. The first morning specimen is preferable.

## XII. **Viral Cultures**

- A. Special collection kits are required and available from the laboratory.
- B. Specimens are collected as required for bacterial (routine) cultures and sent promptly to the lab.

## XIII. **Wound Cultures, Routine Aerobic**

- A. Clean all drainage from wound using gloves and sterile gauze. If the wound is healed over with a scab, this should be removed before collecting the specimen.
- B. The specimen should be collected using a Eswab/Culturette --- (provided to lab clients) taking care not to contaminate the swab with surface organisms around the wound. It may be necessary to use pressure on the wound to cause drainage.
- C. The preferred method of collection is by needle aspiration. Insert a needle attached to a syringe through properly prepped adjacent intact skin to the depth of the wound and aspirate infected material. Transfer material to transport tube tube.

## XIV. **Microbial Stains and Smears**

- A. General --- Only one Eswab is required for all tests selected. If a Culturette is used, a separate Culturette must be submitted for each test ordered.
- B. Gram stain
  - 1. Not recommended for throat, fecal, or blood specimens.
  - 2. Performed routinely on CSF and other body fluids (excluding urine) and expectorated sputum when a routine (bacterial) culture is requested.
  - 3. Performed routinely on wound and surgical specimens when a routine (bacterial) culture is requested and enough material is submitted.
  - 4. Must be requested on other specimens (eg urine, cervical specimens).
  - 5. May be requested separately without a culture.
- C. AFB smear
  - 1. Not performed routinely on fecal, gastric, blood, and non catheterized urine specimens.
  - 2. Performed routinely on other specimens whenever an AFB culture is requested.
  - 3. May be requested separately without a culture.
- D. KOH prep
  - 1. See fungus cultures for proper specimen collection.
  - 2. Not performed on CSF. If requested, an India Ink prep will be substituted.
  - 3. Nor performed on vaginal specimens. If requested a wet prep will be substituted.
- E. India Ink prep -- Performed on CSF when requested.

- F. Wet prep-Collect specimen on a swab and immerse in a small amount of saline. Send to laboratory promptly.
- G. Fecal smear for WBCs -- A pea-sized portion or a few milliliters (if liquid) of feces is placed in an empty fecal transport container).
- H. Trichrome stain -- Performed routinely on all fecal specimens when a comprehensive ova and parasite determination has been requested.

## **Anatomic Pathology and Cytology Specimens**

### **Surgical Pathology Specimens**

Routine specimens are submitted in 10% neutral buffered formalin. Ideally, the formalin to tissue ratio should be 10:1. Large specimens may be submitted fresh. Tissue may be submitted fresh for special studies in an appropriate container.

Some biopsies require special consideration with prior arrangements with Histology. These include:

- Skin biopsies for immunofluorescence: to be submitted in Michelle's solution
- Renal biopsies: to be picked up by Histology personnel at the time of biopsy
- Skeletal muscle biopsies: to be delivered stat in a muscle clamp in a fresh state to Histology
- Breast biopsies when carcinoma may be present: to be placed in formalin and delivered to histology as soon as possible.
- Lymph nodes when there is a suspicion of malignant lymphoma: to be delivered as quickly as possible in a fresh state for possible flow cytometric studies, etc.

All the special handling procedures should be indicated on the requisition.

### **Thin Prep Pap Smear**

- Endocervical Brush/Spatula protocol- Obtain an adequate sampling from the ectocervix using the plastic spatula. Rinse the spatula as quickly as possible into the PreservCyt Solution vial by swirling vigorously in the vial 10 times. Discard the spatula.  
Obtain an adequate sampling from the endocervix using the endocervix brush device. Insert the brush into the cervix until only the bottom most fibers are exposed. Slowly rotate brush  $\frac{1}{4}$  to  $\frac{1}{2}$  turn in one direction. Do not over rotate. Rinse the brush as quickly as possible in the PreservCyt solution by rotating the device in the solution 10 times while pushing against the PreservCyt vial wall. Swirl the brush vigorously to further release material. Discard the Brush.  
Tighten the cap so that the torque line on the cap passes the torque line on the vial.

- Record the patient name and ID number on the vial.
- Broom like device protocol- Obtain an adequate sampling from the cervix using the broom like device. Insert the central bristles of the broom into the endocervical canal deep enough to allow the shorter bristles to fully contact the ectocervix. Push gently and rotate the broom in a counter clockwise direction five times.  
Rinse the broom as quickly as possible into the PreservCyt solution vial by pushing the broom in to the bottom of the vial 10 times, forcing the bristles apart. As a final step, swirl the broom vigorously to further release material. Discard the collection device.  
Tighten the cap so that the torque line on the cap passes the torque line on the vial.  
Record the patient name and ID number on the vial.

**Vaginal smear for hormone evaluation:** Lightly scrape the lateral walls with spatula. Spread material evenly onto slide and fix immediately.

## Medical Cytology

This section refers to all non-Gyn specimens with the exception of fine needle aspirates, sputums, and urines, which are more specifically addressed under other headings. These specimens are of two types: examination of premade smears (brushes from GI tract, ureter, bronchus, touch preps) and preparation of slides from fluids (pleural, peritoneal, cyst fluid, etc).

Glass slides and fixative may be obtained from Cytology. Fluids can be delivered in any appropriate container.

### Collection

- Fluids: Fluids may be refrigerated and delivered to the laboratory as quickly as possible. If a prolonged interval between obtaining the specimen and delivering it to the laboratory is anticipated, cellular detail may be better preserved by adding an equal volume of 50% alcohol. On occasion, special studies such as flow cytometry or electronmicroscopy may be indicated on fluids. Arrangements should be made with the pathologist for these specialized studies prior to obtaining the fluid for cytologic examination.
- Premade smears: Slides may be prepared from bronchial or gastrointestinal brushes by rolling the brush on the slide. It is important not to use excessive pressure, and it is critical that the slide be immediately fixed by immersion in 95% alcohol in order to prevent air drying. Slides must be labeled in pencil on the frosted end with the patient's name and date of birth.

## Fine Needle Aspirate Cytology

The use of a thin needle to aspirate cells means that cells can be aspirated from almost any site in the body. This procedure is less traumatic to the patient while producing samples that may be diagnosed cytologically.

### II. Specimen and Storage:

Any cell sample may be aspirated with a small gauge needle.

### III. Reagents, Materials, and Equipment:

95% ethanol	Syringes
CytoLyt solution	Needles Syringe holder (gun)
Flow media	Disposable alcohol swabs
Betadine swabs	Microscope slides
3 step stain	Microscope
Sterile container	Gauze pads
Pencil	Requisition
Immediate evaluation worksheet	

### IV. Procedure:

- A. Fine needle aspirations of deep sites are performed by clinician under radiographic guidance. Fine needle aspiration of superficial "lumps and bumps" may be performed

by properly trained physicians. Pathologists will perform the procedure upon request of the referring physician. At the request of the Clinician, a Pathologist or Cytotechnologist will provide on-site immediate adequacy evaluation for CT or Ultrasound guided FNA's, brushings, and core biopsies.

- B. Each microscope slide must be labeled with the patient's name and date of birth in pencil on the frosted end, and must be accompanied by the appropriate requisition.
- C. The specimen is obtained by aspiration with 23 or 25-gauge needles in accordance with previous training with special attention given to anatomic considerations and sterile technique. Ideally, 2-3 passes should be performed.
- D. The specimen is expelled in a small drop on a glass slide. Another glass slide is placed on top and the two are gently pressed together and pulled apart, smearing the specimen across the two slides.
- E. Fix one slide by immediately submerging in 95% alcohol. Allow other smear to air dry. Stain with 3 step stain (Diff Quik) if immediate evaluation is requested.
- F. Repeat this procedure if additional smears are desired or if additional passes are made.
- G. If flow cytometry is desired, one pass should be deposited in flow media (RPMI), which is labeled with the patient's name and other unique identifier.
- H. Additional specimen may be deposited in CytoLyt Solution and / or needles should be rinsed into CytoLyt for additional processing. CytoLyt tube must be appropriately labeled with two patient identifiers and the time of collectio

## **Sputum Cytology**

Sputum cytology is a very useful method of cytologic investigation of the respiratory tract. Multiple samples can be obtained without causing any harm or discomfort to the patient, and the diagnostic yield of the cells is excellent.

### **II. Specimen and Storage:**

Sputum collected in the manner described below.

### **III. Reagents, Materials, and Equipment:**

70% ETOH	Identification Label
Collection jar	Cytology requisition form
Safety label	

### **IV. Quality Control:**

- A. If too many inflammatory cells are found, the patient should received antibiotic and expectorant therapy 3 to 5 days, then the sputum should be repeated.
- B. The absence of pulmonary alveolar macrophages containing carbon particles shows the sample to be unsatisfactory, containing only saliva.

### **V. Procedure:**

- A. A cytology requisition, collection jar containing 70% ETOH with an alcohol warning label, should be obtained from cytology.
- B. The requisition should be completed in full, the patient's name written on the collection jar (NOT the lid alone), and the safety label affixed to the jar.

C. If is recommended that a minimum of three different samples be obtained. This is ideally done immediately after the patient awakens early in the morning for three consecutive days. The samples should NOT be collected during the same day.

D. The patient should be instructed to rinse his mouth with water only prior to the procedure.

E. The patient should be instructed to inhale repeatedly to full capacity of his lungs and exhale the air with an explosive cough. The resulting expectorant should be caught in the collection jar. This step should be repeated two or three times.

F. The collection jar should be capped tightly and then shaken briskly to dispense the mucus threads and cells and allow for adequate fixation.

G. It is a good idea to disinfect the outside of the container after each specimen is collected to guard against contamination by pathogens, like tuberculosis.

H. After the sputum series has been collected, the completed requisition and the specimen should be delivered to cytology.

I. Process specimen as outlined elsewhere in this manual. For specimens on inpatients, fresh samples with no fixative are acceptable.

VII. **Limitations:**

a. Due to the usual high cellularity of sputum samples, careful screening of the smear is required.

b. The specimen is NOT limited by degeneration or breakdown if properly fixed and may remain un-refrigerated during the specimen collection period.

c. Specimens MUST be clearly marked with the warning label and patients warned NOT to drink the fixative.

## **Urine Cytology**

Freshly voided or catheterized urine should be refrigerated and promptly forwarded to the Cytology Laboratory. If a delay of more than half a day is anticipated in transport to Cytology, an equal volume of 50% alcohol should be added to the specimen to preserve cellular detail. The specimen should be forwarded in an appropriate container labeled with the patient's name and accompanied by a completed requisition.

## PROFILES AND SURVEYS

(For methodologies, test performance schedules and normal values refer to alphabetical listings of individual tests)

### BASIC METABOLIC PROFILE/ CHEM 8

**Specimen Requirements:** 2 mL serum (Gold )

**Profile Includes:**

BUN	Glucose
Chloride	Potassium
CO2	Sodium
Creatinine	Calcium

### COMPREHENSIVE METABOLIC PROFILE /CHEM 14

**Specimen Requirements:** 2 mL serum (Gold )

**Profile Includes:**

Albumin	Calcium	CO2
Alkaline Phosphatase	Cholesterol	Uric Acid
AST (SGOT)	Glucose	
ALT (SGPT)	LDH	
Bilirubin, Total	Phosphorous	
BUN	Protein, Total	

### HEPATITIS PROFILE, VIRAL - COMPREHENSIVE

**Specimen Requirements:** 2 mL serum (Gold )

**Profile Includes:**

HBS-AG	Hepatitis B Surface Antigen
HBS-AB	Hepatitis B Surface Antibody
HBS-Core	Hepatitis B Core Antibody
HAV-AB - Igm	Hepatitis A Igm Antibody
HCV-AB	Hepatitis C Antibody



### HEPATITIS PROFILE, ACUTE (VIRAL)

**Specimen Requirements:** 2 mL serum (Gold )

**Profile Includes:**

HBS-AG	Hepatitis B Surface Antigen
Core- IgM	Hepatitis B Core IgM
HAVAB-IgM	Hepatitis A Igm Antibody
HCV-AB	Hepatitis C Antibody

### HEPATITIS PROFILE, CHRONIC (VIRAL)

**Specimen Requirements:** 2 mL serum (Gold )

**Profile Includes:**

HBS-AG	Hepatitis B Surface Antigen
HBS-AB	Hepatitis B Surface Antibody
HBS-Core	Hepatitis B Core Antibody
HCV-AB	Hepatitis C Antibody

### LIPID PROFILE

**Specimen Requirements:** 2 mL serum; 12-hour fast required (Gold )

**Profile Includes:**

Calculated LDL	HDL Cholesterol
CHD Risk	Triglycerides
Cholesterol, Total	

### LIPID PROFILE (REFLEX DIRECT LDL-C)

**Specimen Requirements:** 2ml serum ; 12 hour fast required (Gold )

**Profile Includes:**

Same as lipid profile above except that if Triglyceride exceeds 300 or the LDL can not be calculated, then a Direct LDL-Cholesterol is performed.

### LIVER (Hepatic)PROFILE

**Specimen Requirements:** 3 mL serum (Gold )

**Profile Includes:**

Albumin	Bilirubin, Total and Direct
Alkaline Phosphatase	Total Protein
ALT (SGPT)	
AST (SGOT)	

## OBSTETRIC PANEL

**Specimen Requirements:** 6 mL clotted blood (Gold )\*  
7 mL EDTA Whole Blood (7 mL Pink top)\*  
5 mL EDTA whole blood (5 mL lavender)\*  
\* Tubes must be labeled with name, date, time, social security number, and phlebotomist initials.

**Profile Includes:**

CBC with Differential	Hepatitis B Surface antibody
Rubella Antibody	RPR
Blood Type and Antibody Screen	HIV Antibody

## PRENATAL PROFILE I

**Specimen Requirements:** 6 mL clotted blood (Gold )\*  
7 mL EDTA Whole Blood (7 mL Pink top)\*  
5 mL EDTA whole blood (5 mL lavender)\*  
\* Tubes must be labeled with name, date, time, social security number, and phlebotomist initials.

**Profile Includes:**

Antibody Screen	Pap Test
Anti-HBsAG	RPR
Blood Type and Rh	Rubella
CBC with Differential	

## PRENATAL PROFILE II

**Specimen Requirements:** 6 mL clotted blood (Gold )\*  
7 mL EDTA Whole Blood (7 mL Pink top)\*  
5 mL EDTA whole blood (5 mL lavender)\*  
\* Tubes must be labeled with name, date, time, social security number, and phlebotomist initials.

**Profile Includes:**

Antibody Screen	Pap Test
Anti-HbsAg	RPR
Blood Type and Rh	Rubella
CBC with Differential	Sickle Cell Test

## RENAL FUNCTION PANEL

**Specimen Requirements:** 3 mL Serum (Gold )

**Profile Includes:**

Albumin	Glucose
Calcium	Phosphorus
Electrolytes	Bun
Creatinine	

